ADAPTATIONS OF THE PIAVA (Leporinus obtusidens) JUVENILE EXPOSED TO HYPOXIA*

Jhon Edison JIMENEZ¹ and Evoy ZANIBONI-FILHO²

ABSTRACT

The piava (*Leporinus obtusidens*) is a migratory freshwater fish. There is relatively little information on the hematological and morphological adaptations of tropical freshwater fish when subjected to chronic and sublethal concentrations of dissolved oxygen (DO). The aim of this study was to verify the existence of adaptations of the gills and blood in juvenile piava exposed to different concentrations of DO. The juveniles used in this experiment were cultured for 70 days at different DO concentrations (1.74 ± 0.04 , 3.54 ± 0.06 , 5.34 ± 0.05 and 7.34 ± 0.02 mg L⁻¹) and kept at mean water temperature of 26.4°C. Individuals subjected to conditions of severe hypoxia (1.74 mg L^{-1} DO) showed a significant increase in the number of erythrocytes (P<0.05) and a decrease in erythrocyte corpuscular volume (P<0.05), but no changes were observed in gill lamellae. This study concludes that *L. obtusidens* juveniles undergo hematological change when exposed to conditions of severe hypoxia.

Keywords: Dissolved oxygen; hematocrit; erythrocytes; gills; blood

ADAPTAÇÕES DE JUVENIS DE PIAVA (Leporinus obtusidens) EXPOSTOS À HIPÓXIA

RESUMO

A piava (*Leporinus obtusidens*) é um peixe migrador de água doce. São relativamente escassas as informações de adaptações hematológicas e/ou morfológicas apresentadas por peixes tropicais de água doce quando submetidos à concentrações crônicas e subletais de oxigênio dissolvido (OD). O objetivo deste trabalho foi verificar a existência de adaptações branquiais e sanguíneas em juvenis de piava submetidas a diferentes concentrações de oxigênio dissolvido. Para tal, juvenis foram cultivados durante 70 dias em diferentes concentrações de oxigênio dissolvido (1,74 ± 0,04, 3,54 ± 0,06, 5,34 ± 0,05 e 7,34 ± 0,02 mg L⁻¹) a uma temperatura média da água de 26,4°C. Indivíduos submetidos às condições de hipóxia severa (1,74 mg L⁻¹ de OD) apresentaram aumento significativo do número de eritrócitos (*P*<0,05) e redução no volume corpuscular dessas células (*P*<0,05). Não foram observadas modificações morfológicas nas lamelas branquiais. É possível concluir que juvenis de *L. obtusidens* apresentam adaptações hematológicas quando submetidos à condição de hipóxia severa.

Palavras chave: Oxigênio dissolvido; hematócrito; eritrócitos; brânquias; sangue

Nota Científica: Recebida em 23/02/2013 - Aprovada em 16/08/2013

¹ Universidade Federal de Santa Catarina (UFSC). e-mail: jejimenezro@gmail.com (corresponding author)

² Departamento de Aquicultura - Universidade Federal de Santa Catarina (UFSC). e-mail: zaniboni@cca.ufsc.br

Address: Laboratório de Biologia e Cultivo de Peixes de Água Doce (LAPAD), Departamento de Aquicultura, Universidade Federal de Santa Catarina. Rodovia SC 406, nº 3532 – Florianópolis – CEP: 88066-000 – SC – Brazil

reaerai de Santa Catarina. Roadona SC 400, nº 5552 - Floranopous - CEF. 66000-000 - SC * Financial company. Tracatal al Financia, FADECC and CND-

^{*} Financial support: Tractebel Energy, FAPESC and CNPq

INTRODUCTION

Hematological parameters are a useful tool for understanding some of the physiological strategies used by the fish when they are exposed to hypoxia conditions.

An increase in the hematocrit and the number of erythrocytes (red blood cells - RBC) are some of the classic changes observed in fish subjected hypoxia (VALENZUELA et al., 2002; to BALDISSEROTTO et al., 2008; BALDISSEROTTO, 2009). Increases in RBC are due to the fact that most of the oxygen transported in the blood is attached to hemoglobin encapsulated inside these cells (BRAUNER and VAL, 2006). The spleen may also promote a rapid response to hypoxia conditions by contributing with additional RBC (VALENZUELA et al., 2002). Some species of tropical fish can adapt to local conditions of low dissolved oxygen (DO) levels via morphological modifications of the gills; an increase of respiratory surface area allows for higher oxygen uptake (CHAPMAN and HULEN, 2001; CHAPMAN et al., 2002, 2008; CRAMPTON et al., 2007; ONG et al., 2007).

Leporinus obtusidens (Anostomidae) is a large migratory freshwater fish that can reach weights exceeding 7.5 kg (TAITSON *et al.*, 2008) and is found in the South American basins of the Paraná, São Francisco and Uruguay rivers (OLDANI and OLIVEROS, 1984; TABLADO *et al.*, 1988; SATO *et al.*, 2000; ZANIBONI-FILHO and SCHULZ, 2003). Leporinus obtusidens is an omnivorous species (HAHN *et al.*, 1998) and it is of great commercial importance for the quality and flavor of its meat.

The relative lack of information about the response of South American freshwater species to different levels of DO hinders the establishment of criteria for adequate cultivation conditions. It is known that prolonged exposure to stressful levels of DO can affect the physiological processes of fish, interfering with feeding rates, and limiting growth (KARIM *et al.*, 2002; WILHELM FILHO *et al.*, 2005) and reproduction (DABROWSKI *et al.*, 2003).

This study aims to assess whether hematological and morphological adjustments in

juvenile *L. obtusidens* are made in response to different concentrations of DO.

MATERIALS AND METHODS

Piava juveniles (*L. obtusidens*) were obtained from the hormonal induction of the wild breeding stock from LAPAD/UFSC, captured in the Upper Uruguay River basin and maintained and bred in captivity. The juvenile piava (9.22 ± 1.76 g and 10.19 ± 0.64 cm) were exposed to four different concentrations of DO: 1.74 ± 0.04 , 3.54 ± 0.06 , 5.34 ± 0.05 and 7.34 ± 0.02 mg L⁻¹; tested in triplicate. These values are equivalent to 21, 47, 70 and 92% of the oxygen saturation, respectively.

Seventy-liter fiberglass tanks were stocked with 20 fish per tank. The fish were reared over 70 days of experimentation and kept at an average temperature of 26.4°C. Piava juveniles were fed twice daily (08:00 and 16:00 hours) with commercial feed (42% crude protein (CP) and 3,800 kcal kg⁻¹ of digestible energy) until apparent satiation during almost 10 minutes. To avoid gas exchange between atmospheric air and water, each tank water surface were covered with plastic. Survival was evaluated at the end of each day. The fish were cultivated in a closed recirculating system, using mild saline water (NaCl at 1 ppt), and water was renewed 250% per day in each fiberglass tanks. Dissolved oxygen concentrations maintained through aeration were and incorporation of nitrogen gas.

Dissolved oxygen were monitored every hour from 08:00 to 22:00 hours (15 times per day); temperature were monitored three times per day (08:00, 12:00 and 16:00 hours), and pH and conductivity were measured twice daily (08:00 and 16:00 hours). Measurements of oxygen and temperature were taken using YSI model 550A (Yellow Springs Instruments Company, OH, USA), and pH and conductivity were evaluated using a YSI model 63 multiparameter probe (Yellow Springs Instruments Company, OH, USA). Hardness and concentrations of total ammonia, nitrite and alkalinity were evaluated once a week using the colorimetric titration method (Alpha Tecnoquímica, SC, Brazil). All the water quality parameters were taken from the center of the tank between the inlet and outlet of water.

At the end of 70 days of cultivation, five fish from each tank were randomly selected and anesthetized with 100 mg L⁻¹ of eugenol. These fish were individually weighed, and then 1 mL of blood was collected with the aid of needles and syringes with EDTA, via caudal puncture.

The material collected was used to determine the hematocrit (Ht), obtained through the micro hematocrit technique by centrifuging the blood at 2,800 rpm for five minutes. Prior to blood dilution in a 1:100 saline solution, the number of RBC from this same sample was counted in a Neubauer chamber. The mean corpuscular volume (MCV = Ht x 10/RBC) was then calculated. The protocol used to handle fish was previously approved by ethics committee (CEUA-UFSC) by number PP00788. The fish were sacrificed with an overdose of eugenol (300 mg L-1), to determine the spleen-somatic index (SSI = spleen weight x 100/fish weight) using an electronic balance (0,001 g). After 24 h of immersion in 10% phosphate-buffered formalin fixative, the second left gill arch was cut away and then routinely processed for paraffin embedding. The gill arches were serially sectioned in 4 µm increments and then stained with hematoxylin and eosin. The slides were viewed using a LEICA DM3000LED light microscope with ocular lens ruler (10 mm) to determine the total length (L) (measured from the edge adjacent to the filament to the most distal point of the lamellae from the filament) and width (W) (measured parallel to the filament at the base of the lamellae from one edge to the other) of the lamellae. Measurements were obtained using the methodology described by ONG *et al.* (2007).

The variables Ht, RBC, MCV, SSI, L and W were subjected to an analysis of variance (ANOVA), followed by the Tukey test (P<0.05). All analyses were performed using version 8.0 of the statistical program SAS (SAS Institute Inc., NC, USA).

RESULTS

Survival rates were 100% for all treatments. The mean values of Ht, L and W were similar (P>0.05) for fish kept at different DO concentrations. The highest RBC and the lowest MCV were observed in fish exposed to severe hypoxia (1.74 mg L⁻¹) (P<0.05). The SSI values were lower in fish kept in severe hypoxia (1.74 mg L⁻¹) than in those kept at 3.54 mg L⁻¹ DO, although similar values were observed in fish kept in normoxia (Table 1).

The water quality parameters were similar in all treatments, showing the following mean values (\pm SE): temperature: 26.44 \pm 0.30°C; pH: 6.47 \pm 0.19; electrical conductivity: 1,821.17 \pm 89.79 μ S; total ammonia: 0.40 \pm 0.08 mg L⁻¹; nitrite: 0.44 \pm 0.12 mg L⁻¹; hardness: 102.67 \pm 2.51 mg L⁻¹ CaCO₃; alkalinity: 18.49 \pm 1.26 mg L⁻¹ CaCO₃.

Table 1. Mean values (± SE) of hematological parameters and morphological gill characteristics in *Leporinus obtusidens* submitted for 70 days at different oxygen concentrations.

Dissolved oxygen (mg L ⁻¹) [%]	Ht (%)	RBC (10 ⁵ μL ⁻¹)	MCV (μ ³)	SSI (%)	L (µm)	W (μm)
(1.74±0.04) [21.50±1.34]	31.54 ± 7.33a	6.78 ± 0.95a	$46.11 \pm 4.50b$	$0.019\pm0.001\mathrm{b}$	64.79 ± 14.77a	10.06 ± 1.12a
(3.54±0.06) [47.43±0.67]	31.79 ± 6.57a	$4.87\pm0.07\mathrm{b}$	65.35 ± 13.75ab	$0.027 \pm 0.002a$	54.29 ± 7.81a	10.70 ± 1.95a
(5.34±0.05) [70.35±0.33]	33.47 ± 1.88a	$4.48\pm0.09\mathrm{b}$	$74.68 \pm 4.34a$	$0.024 \pm 0.002ab$	61.95 ± 8.07a	11.79 ± 1.18a
(7.34±0.02) [91.77±1.19]	34.99 ± 3.22a	$5.05 \pm 0.17b$	69.41 ± 8.63a	0.023 ± 0.004 ab	61.75 ± 1.55a	11.67 ± 2.12a

Ht = hematocrit; RBC = number of erythrocytes; MCV = mean corpuscular volume; SSI = spleen somatic index; L = total length of the lamellae; W = width of the lamellae. Different letters in the same column indicate significant differences as determined by the Tukey test. (P<0.05).

DISCUSSION

Leporinus obtusidens juveniles undergo a hematological adaptation to hypoxia by

increasing their RBC numbers when subjected to low DO concentrations (1.74 mg L⁻¹). This same response to hypoxia has been observed in other fish species, such as *Astronotus ocellatus* (MUUSZE *et al.*, 1998), *Piaractus mesopotamicus* (GÓMEZ-MANRIQUE *et al.*, 2009), *Astatoreochromis alluaudi* and *Haplochromis ishmaeli* (RUTJES *et al.*, 2007). VALENZUELA *et al.*, (2002) also showed a reduction in spleen size associated with an increased number of RBC in *Oncorhynchus mykiss*. In this study, we observed a reduction of the SSI and an increased number of RBC for fish kept in severe hypoxia.

As a result of the increased number of RBC, the hematocrit is expected to increase, or the MCV is expected to decrease. Some authors suggest that a reduction of RBC volume favors rapid oxygenation of hemoglobin. This oxygenation is a result of an increased relationship between the surface area and volume of erythrocytes, thus improving their ability to transport oxygen (WELLS *et al.*, 2005; TAVARES-DIAS *et al.*, 2008). In this study, hematocrit values remained similar in juvenile *L. obtusidens* exposed to different concentrations of oxygen. However, MCV showed a reduction as a result of varying the concentration of DO.

Some fish species showed a morphological remodeling of the gills in response to hypoxia; *Gymnocypris przewalskii* demonstrated a gradual lengthening of lamellae when subjected to values of 0.3 mg L⁻¹ DO for 24 hours (MATEY *et al.*, 2008), and *Pseudocrenilabrus multicolor victoria* demonstrated a greater average gill filament length when kept for a minimum of one year at a concentration of 1.3 mg L⁻¹ DO (CHAPMAN *et al.*, 2008).

Leporinus obtusidens juveniles showed no morphological changes to total length and width of the gill lamellae in response to reductions in oxygen values. It is possible that the hypoxia intensity and exposure time used in this study were not sufficient for the species to manifest any such adaptive response.

According to NILSSON (2007), the apoptosis and mitosis mechanisms in morphological remodeling of the gills represent high energy costs.

Morphological adaptation would therefore be less used by species facing hypoxic conditions for short periods of time. Additionally, an increase in respiratory surface area allows for greater uptake of oxygen and, also would increase the flow of ions and water through the gills. These augmentations result in additional osmoregulation costs and make fish more vulnerable to pathogens and toxic substances (SOLLID and NILSSON, 2006).

CONCLUSION

Leporinus obtusidens juveniles showed hematological adaptations to hypoxia, with an increased number and reduced volume of RBC when oxygen concentrations were maintained at 1.74 mg L⁻¹. When the fish were kept in water with oxygen concentrations equal to or higher than 3.54 mg L⁻¹, they showed no hematological changes. These results may suggest a condition of comfort for *L. obtusidens*.

ACKNOWLEDGMENTS

We would also like to thank the technical team at LAPAD for their support during the study.

REFERENCES

- BALDISSEROTTO, B. 2009 *Fisiologia de peixes aplicada à piscicultura*. Second ed. SantaMaria: Editora da UFSM. 349p.
- BALDISSEROTTO, B.; CHIPPARI-GOMES, A.R.; LOPES, N.P.; BICUDO, J.E.P.W.; PAULA-SILVA, M.N.; ALMEIDA-VAL, V.M.F.; VAL, A.L. 2008 Ion fluxes and hematological parameters of two teleosts from the Rio Negro, Amazon, exposed to hypoxia. *Brazilian Journal of Biology*, 68(3): 571-575.
- BRAUNER, C.L. and VAL, A.L. 2006 Oxygen transfer. In: VAL, A.L.; ALMEIDA-VAL, V.M.F.; RANDALL, D.J. *The physiology of tropical fishes*. Amsterdam: Elsevier Science Publishers. p.277-306.
- CHAPMAN, L.J. and HULEN, K.G. 2001 Implications of hypoxia for the brain size and gill morphometry of mormyrid fishes. *Journal of Zoology*, 254: 461-72.
- CHAPMAN, L.J.; ALBERT, J.; GALIS, F. 2008 Developmental plasticity, genetic differentiation, and hypoxia-induced trade-offs in an African cichlid fish. *The Open Evolution Journal*, 2: 75-88.
- CHAPMAN, L.J.; CHAPMAN, C.A.; NORDLIE, F.G.; ROSENBERGER, A.E. 2002 Physiological

refugia: swamps, hypoxia tolerance and maintenance of fish diversity in the Lake Victoria region. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 133: 421-437.

- CRAMPTON, W.G.R.; CHAPMAN, L.J.; BELL, J. 2007 Interspecific variation in gill size is correlated to ambient dissolved oxygen in the Amazonian electric fish *Brachyhypopomus* (Gymnotiformes: Hypopomidae). *Environmental Biology of Fish*, 83: 223-235.
- DABROWSKI, K.; RINCHARD, J.; OTTOBRE, J.S.; ALCANTARA, F.; PADILLA, P.; CIERESZKO, A.; DE JESUS, M.J.; KOHLER, C.C. 2003 Effect of oxygen saturation in water on reproductive performances of pacu *Piaractus brachypomus*. *Journal of the World Aquaculture Society*, 34(4): 441-449.
- GÓMEZ-MANRIQUE, W.; MASSAGO, H.; ABREU-SANTOS, D.J.; CRISCUOLO-URBINATI, E. 2009 Respuesta del *Piaractus mesopotamicus* a estímulos de persecución e hipoxia. *Revista Orinoquia*, 13(2): 93-100.
- HAHN, N.S.; AGOSTINHO, A.A.; GOMES, L.C.; BINI, L.M. 1998 Estrutura trófica da ictiofauna do reservatório de Itaipu (Paraná-Brasil) nos primeiros anos de sua formação. *Interciência*, 23(5): 299-305.
- KARIM, M.R.; SEKINE, M.; UKITA, M. 2002 Simulation of eutrophication and associated occurrence of hypoxic and anoxic condition in coastal bay in Japan. *Marine Pollution Bulletin*, 45: 280-285.
- MATEY, V.; RICHARDS, J.G.; WANG, Y.; WOOD, C.M., ROGERS, J.; DAVIES, R.; MURRAY, B.W.; CHEN, X.Q.; DU, J.; BRAUNER, C.J. 2008 The effect of hypoxia on gill morphology and ionoregulatory status in the Lake Qinghai scaleless carp, *Gymnocypris przewalskii. The Journal of Experimental Biology*, 211: 1063-1074.
- MUUSZE, B.; MARCON, J.; VAN DEN THILLART, G.; ALMEIDA-VAL, V. 1998 Hypoxia tolerance of Amazon fish respirometry and energy metabolism of the cichlid Astronotus ocellatus. Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology, 120: 151-156.

- NILSSON, G.E. 2007 Gill remodeling in fish a new fashion or an ancient secret?. *The Journal of Experimental Biology*, 210: 2403-2409.
- OLDANI, N.O. and OLIVEROS, O.B. 1984 Estudios limnológicos en una sección transversal del tramo medio del río Paraná. XII. Dinámica temporal de peces de importancia económica *Revista de la Asociación de Ciencias Naturales del Litoral*, 15(2): 175-183.
- ONG, K.J.; STEVENS, E.D.; WRIGHT, P.A. 2007 Gill morphology of the mangrove killifish (*Kryptolebias marmoratus*) is plastic and changes in response to terrestrial air exposure. *The Journal of Experimental Biology*, 210: 1109-1115.
- RUTJES, H.A.; NIEVEEN, M.C.; WEBER, R.E.; WITTE, F.; VAN DEN THILLART, G.E.E.J.M. 2007 Multiple strategies of Lake Victoria cichlids to cope with lifelong hypoxia include hemoglobin switching. American Journal of Physiology - Regulatory, Integrative and Comparative Physiology, 293: R1376-R1383.
- SATO, Y.; FENERICH-VERANI, N.; VERANI, J.R.; VIEIRA, L.J. S.; GODINHO, H.P. 2000 Induced reproductive responses of the neotropical anostomid fish *Leporinus elongatus* Val. under captive breeding. *Aquaculture Research*, 31: 189-193.
- SOLLID, J. and NILSSON, G.E. 2006 Plasticity of respiratory structures – Adaptive remodeling of fish gills induced by ambient oxygen and temperature. *Respiratory Physiology & Neurobiology*, 154: 241-251.
- TABLADO, A.; OLDANI, N.O.; ULIBARRIE, L.; PIGNALBERI DE. HASSAN, C. 1988 Cambios estacionales de la densidad de peces en una laguna del valle aluvial del río Paraná (Argentina). *Revue d'Hydrobiolgie Tropicale*, 21(4): 335-348.
- TAITSON, P.F.; CHAMI, E.; GODINHO, H.P. 2008 Gene banking of the neotropical fish *Leporinus obtusidens* (Valenciennes, 1836): A protocol to freeze its sperm in the field. *Animal Reproduction Science*, 105: 283-291.
- TAVARES-DIAS, M.; MORAES, F.R.; IMOTO, M.E. 2008 Hematological parameters in two neotropical freshwater teleost, *Leporinus macrocephalus* (Anostomidae) and *Prochilodus*

lineatus (Prochilodontidae). *Bioscience Journal*, 24(3): 96-101.

- VALENZUELA, A.; ALVEAL, K.; TARIFEÑO, E. 2002 Respuestas hematologicas de truchas (Oncorhynchus mykiss, Walbaum 1792) a estres hipoxico agudo: serie roja. Gayana, 66(2): 255-261.
- WELLS, R.M.G.; BALDWIN, J.; SEYMOUR, R.S.; CHRISTIAN, K.; BRITTAIN, T. 2005 Red blood cell function and haematology in two tropical freshwater fishes from Australia. *Comparative Biochemistry and Physiology Part A: Molecular & Integrative Physiology*, 141: 87-93.
- WILHELM FILHO, D.; TORRES, M.A.; ZANIBONI-FILHO, E.; PEDROSA, R.C. 2005 Effect of different oxygen tensions on weight gain, feed conversion, and antioxidant status in piapara, *Leporinus elongatus* (Valenciennes, 1847). *Aquaculture*, 244: 349-357.
- ZANIBONI-FILHO, E. and SCHULZ, U.H. 2003 Migratory fishes of the Uruguay river. In: CARAOLSFELD, J.; HARVEY, B.; ROSS, C.; BAER, A. Migratory fishes of south America: biology, social importance and conservation status. Victoria: World Fisheries Trust/IDRC/World Bank. p.157-194.