

BOLETIM DO INSTITUTO DE PESCA

ISSN 1678-2305 online version Scientific Note

USE OF 17β-ESTRADIOL FOR *Leporinus macrocephalus* FEMINIZATION*

ABSTRACT

Thiago Scremin Boscolo PEREIRA^{1,2} Camila Nomura Pereira BOSCOLO¹ Sergio Ricardo BATLOUNI^{3*} (D

¹Centro Universitário de Rio Preto – UNIRP, Rua Ivete Gabriel Atique, 45, CEP: 15025-400, São José do Rio Preto, SP, Brazil.

²FACERES, Avenida Anísio Haddad, 6751, CEP: 15090-305, São José do Rio Preto, SP, Brazil.

^{3*}Universidade Estadual Paulista – UNESP, Centro de Aquicultura da UNESP – CAUNESP, Via de Acesso Professor Paulo Donato Castellane, s/n, CEP: 14884-900, Jaboticabal, SP, Brazil. sergio.ricardo@unesp.br (corresponding author)

*This study was financed in part by the Fundacão de Amparo à Pesquisa do Estado de São Paulo - FAPESP (FAPESP 2010/08334-7 - doctoral scholarship).

Received: September 23, 2019 Accepted: July 02, 2020 The aim of this study was to evaluate the effects of the use of a diet supplemented with 50 or 100 mg kg⁻¹ 17 β -estradiol (E₂) for *Leporinus macrocephalus* feminization. Thus, one hundred and fifty fingerlings with 50 days old post-hatch were randomly distributed in fifteen experimental tanks of 90 L and fed for 60 days on a diet supplemented with 50 or 100 mg kg⁻¹ of E₂. At the end of the experiment, sex ratios were determined through histological and macroscopic observations. Histologically, the differentiated ovaries were evidenced by the presence of numerous nests of oogonia and oocytes in primary growth stage. The female ratio (77%) for the group treated with 100 mg kg⁻¹ E₂ was significantly higher than those of control (52%) and 50 mg kg⁻¹ treatment (48%) groups. These results obtained in this initial study indicated that 100 mg kg⁻¹ of E₂, administered over 60 days, was the most effective treatment for 50 days old *L. macrocephalus* post larval feminization. However, future studies with variations in the application range can bring even better results.

Keywords: sex inversion; monosex fish populations; aquaculture-species; native fish.

UTILIZAÇÃO DE 17β-ESTRADIOL PARA FEMINIZAÇÃO DE Leporinus macrocephalus

RESUMO

O objetivo do estudo foi avaliar o efeito do 17β-estradiol (E_2) na feminização de *Leporinus macrocephalus*. Dessa forma, 150 alevinos com 50 dias de idade foram distribuídos aleatoriamente em 15 tanques experimentais de 90 L e alimentados por 60 dias com dieta suplementada com 50 ou 100 mg kg⁻¹ de E_2 . No final do experimento, as proporções sexuais foram determinadas por meio de observações histológicas e macroscópicas. Histologicamente, os ovários diferenciados foram evidenciados pela presença de numerosos ninhos de oogonia e oócitos em fase crescimento primário. O percentual de fêmeas (77%) do grupo tratado com 100 mg kg⁻¹ (48%). Os resultados obtidos neste estudo inicial indicaram que 100 mg kg⁻¹ de E_2 , administrados durante 60 dias, foi o tratamento mais efetivo na feminização de *L. macrocephalus* com 50 dias de idade. No entanto, estudos futuros com variações no intervalo de aplicações podem trazer ainda melhores resultados

Palavras-chave: inversão sexual; populações de peixes monosexo; espécies aquícolas; peixes nativos.

INTRODUCTION

One of the major challenges of Brazilian aquaculture is the lack of technological packages for the creation of important native species of fish. According to the yearbook of Brazilian fish farming, there was a drop in native fish production, especially due to the lack of technological packages development (Peixe BR, 2019). The production of monosex fish populations is one of the alternatives that can help to boost the cultivation of native fish (Fernandino and Hattori, 2019). Monosex batches provide high economic profitability in production, adding value to the exclusive production of the sex with the best growth rates (Singh, 2013; Budd et al., 2015; Thuong et al., 2017; Alcántar-Vázquez, 2018; Vidal-López et al., 2019). Among the native species of fish considered of great commercial potential, the *Leporinus macrocephalus* deserves to be

highlighted, since it is a large-sized fish, commercially relevant for fisheries and aquaculture (Pereira et al., 2017). This fish is endemic of the Paraguay River basin and it is a valuable species in aquaculture programs (Morelli et al., 2007; Hashimoto et al., 2010; Muñoz et al., 2011). This species was, according to official fish production data, one of the ten most produced native fish in Brazil in 2018 (IBGE, 2018), mainly due to its high-quality meat (Duke Energy International-Geração Paranapanema S/A., 2003) and acceptance for commercial, subsistence, and sport fishing purposes (Petrere Junior et al., 2002; Giamas and Vermulm Junior, 2004). *Leporinus macrocephalus* is considered of great commercial potential because it presents fast growth in the initial stages, rusticity to the handling and resistance to the temperature variations (Soares et al., 2000; Riffel et al., 2012; Capodifoglio et al., 2015).

Leporinus macrocephalus females show higher growth rates than males (Reidel et al., 2004), which is a factor that can be used to increase productivity without the need to enlarge the area of cultivation, and, in addition, to reduce the time of slaughter. The use of endocrine techniques for sexual inversion is widely used in fish production, and it directs the formation of monosex populations for the genus that presents zootechnical advantages (Devlin and Nagahama, 2002; Cnaani and Levavi-Sivan, 2009; Singh, 2013; Örn et al., 2016; Thuong et al., 2017; Alcántar-Vázquez, 2018; Vidal-López et al., 2019). In aquaculture this may represent significant economic gain, adding value to the product if only individuals from the sex with the best growth rates are produced. Thus, in order to foster fish farming industry, it is necessary to develop technological packages for the creation of native species considered of great commercial potential. However, there are few studies in the literature that carry out the production of monosex population of freshwater tropical native species. Thus, this study aimed to evaluate the effects of the use of a diet supplemented with 50 or 100 mg kg⁻¹ of 17β-estradiol (E₂) for Leporinus macrocephalus feminization.

MATERIAL AND METHODS

Ethical note

This study was conducted in agreement with the precepts of the National Council for the Control of Animal Experimentation (CONCEA) and was approved by the Animal Ethics and Welfare Committee from UNESP, Jaboticabal, SP, Brazil, under permission number 015279/10.

Fish culture

Fifty days old fingerlings obtained from a fishing farm were transferred to the Fish Reproduction Laboratory at the Aquaculture Center of Unesp – CAUNESP, and kept in 500 L indoor stock-tanks (ca. 1 fish 5 L⁻¹) during 10 days for acclimation before the experiment. During acclimatization, the animals were manually fed a pelleted balanced commercial diet (moisture content (max) 10.0%; crude protein (min) 32.0%; ethereal extract (min) 10.0%; fibrous matter (max) 7.0%; ash (max) 10.0%; calcium (max)

1.2%; phosphorus (min) 1.2%) corresponding to 3.0% of total body weight four a day. External biological filters and constant aeration ensured water quality.

Experimental Protocols

After the acclimation period, 150 fingerlings $(4.35 \pm 0.92 \text{ cm})$ 2.12 ± 1.36 g) were randomly distributed in 15 experimental tanks of 90 L in a density of about 1 fish/9 L. A completely randomized design was applied with an equal number of replicates per treatment (N = 5). Experimental diets were enriched with two different concentrations (50 and 100 mg kg⁻¹) of 17β -estradiol (E₂) (Sigma Co., USA). To incorporate E, into manufactured feed, we used a well-established method of spraying ethanol-dissolved steroids onto the food (Lin et al., 2012; Singh, 2013). This procedure involved making stock solutions (1 ppt) of E₂ dissolved in 100% ethanol which was added by spraying to ensure uniform distribution of the hormone. Then the feed was distributed in travs under the light, for 48h for the complete evaporation of the ethanol. Control diets were prepared in the same way, but without the addition of steroid. The fish were fed to satiation, by hand, four times daily throughout the experiment, with feeding times spread over a 12 h period. External biological filters and constant aeration ensured water quality. Water temperature was kept at $27 \pm 1^{\circ}$ C and photoperiod was 12L: 12D. The leftover food and feces were removed weekly, by the drain in the tanks.

Sex determination and histological examination

Sixty days after the onset of the treatments, two fish per replicate were randomly selected and euthanized with a lethal dose of benzocaine (2 g L⁻¹). Biometric analyses were performed by determination of the individual total weight (g) and standard length (cm). Food intake was calculated by recording the consumption in a period of 24 hours. Its calculation was obtained by the daily subtraction between the food quota offered and the leftovers from the subsequent day (Pereira et al., 2015). To determine the sexual differentiation, the gonads were photographed and analysed by stereomicroscope. However, for the definitive diagnosis of the sex, we collected gonad samples from all animals for histological evaluation, aiming to verify the presence of germ and somatic cells in presumptive testicles and ovaries. The cranial, medial, and tail regions of the gonad's tissues were fixed in Bouin solution for routine histological procedures, embedded in Historesin[™] for histological preparation, and stained with hematoxylin-floxin. The changes in sex ratios were determined by light microscopy. The gonadal tissues were classified as undifferentiated and differentiated. For the sex characterization, the following criteria were adopted: for males, the presence of spermatogonia cysts and spermatocytes were sought, and for females, the presence of oocytes in primary growth. Undifferentiated gonadal tissues were characterized by the presence of gonocytes only (detectable by light microscopy). The effects of E₂ on gonadal differentiation were estimated considering the sexual proportions between the treatments.

Statistical analysis

Data normality was verified using the Cramer-von Mises test. Homoscedasticity was checked through the Fmax test. The food intake and body weight were analyzed by comparing different treatments with a one-way analysis of variance (ANOVA). The Chi - square test (χ^2) was used to analyze gonadal differentiation between treatments. A threshold of P≤0.05 was set to infer statistical significance.

RESULTS

Growth and survival

We did not observe significant differences in the body weight of the animals among groups. Mortality was not observed in any of the experimental groups. There was no difference for food intake among the treatments and controls (P = 0.45; Table 1).

Sex ratio and macroscopic evaluation of gonads

Animals treated with 100 mg kg⁻¹ E_2 showed significantly higher female ratio (77.78%) than control (57.14%) and 50 mg kg⁻¹ treatment

(62.50%) (χ^2 , P = 0.01, Table 1). No male animals were observed in any group up to the end of the experimental period. Undifferentiated gonads, found mainly in the control treatment, were constituted by a pair of translucent, homogeneous and elongated structures, located in the dorsal region of the celomatic cavity, parallel to the swim bladder (Figure 1A). On the other hand, ovaries, which were frequent in the treatment 100 mg kg⁻¹ E₂, presented a thicker aspect in the cranial region in relation to the caudal one. In addition, the its coloration changed from translucent to slightly brownish-gray, with the presence of blood vessels (Figure 1B).

Table 1. Average values (\pm standard error) of food intake, body weight and sex ratio (female) of fingerlings fed with different concentrations (50 and 100 mg kg⁻¹) of E₂.

Parameters	Treatments		
	Control	$E_{2}(50 \mathrm{mg kg^{-1}})$	$E_{2}(100 \mathrm{mg kg^{-1}})$
Food intake (g)	15.04 ± 0.80	14.27 ± 0.43	14.01 ± 0.91
Body weight (g)	21.53 ± 1.08	21.26 ± 0.38	21.33 ± 0.74
Female ratio (%)	$57.14\pm4.7~^{\rm b}$	$62.50\pm2.6^{\rm b}$	$77.78 \pm 1.8^{\rm a}$

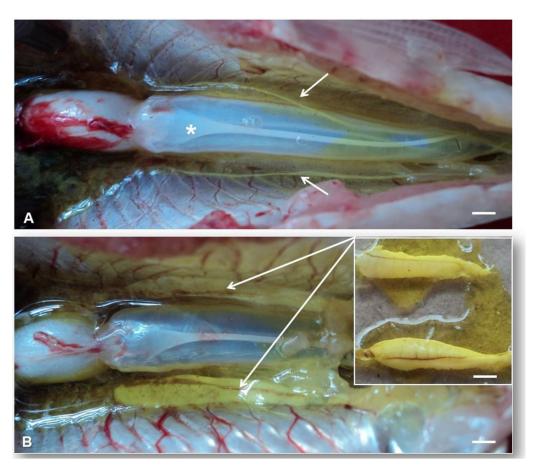


Figure 1. (A) Photograph of undifferentiated gonads observed in the control treatment. The gonads (arrows) were located in the dorsal region of the celomatic cavity, parallel to the swim bladder (asterisk). (B) Photograph of the ovary of *Leporinus macrocephalus* fed with 100 mg kg⁻¹ of E2. The ovaries showed small blood capillaries throughout the gonadal tissue (arrows). The inset shows details of ovaries extracted from the fish for histological processing. Scale bar = 400 μ M.

Histological evaluation of gonads

Undifferentiated gonads were formed by gonocytes and somatic cells. Somatic cells located close or around gonocytes were had elongated to cubic shape. Their basophilic nuclei shape varied according to cell shape. Gonocytes showed relatively large nuclei, which contained a prominent nucleolus and loose chromatin (Figure 2A). Ovaries were evidenced by the presence of numerous nests of oogonia distributed throughout the germinal epithelium. In addition, ovaries with numerous oocytes in primary growth stage were found (Figure 2B).

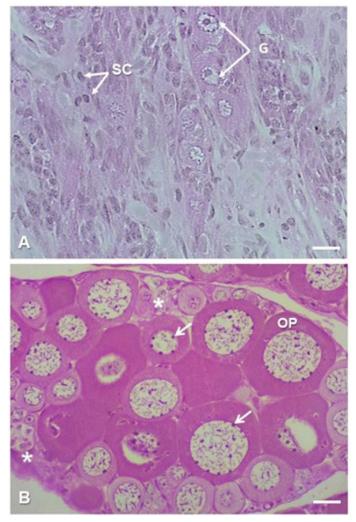


Figure 2. (A) Photomicrographs of cross sections of undifferentiated gonads showing the presence of gonocytes (G) and somatic cells (SC). (B) Photomicrographs of cross sections of the ovary of *Leporinus macrocephalus* fed with 100 mg kg⁻¹ of E_2 showing the presence nests of oogonia (asterisk) and numerous previtellogenic oocytes (OP) in primary growth stage. In this stage multiple nucleoli are displaced to the periphery of the germinal vesicle (arrows). Hematoxylin–floxin. Scale bar: Fig 2A = 20 μ M; Fig 2B = 40 μ M.

DISCUSSION

Leporinus macrocephalus fed with diets supplemented with 100 mg kg⁻¹ of E_2 had significantly higher female ratios, being an effective alternative to promote the process of feminization in this species.

Although sexual steroids have a direct influence on the growth and survival of teleosts (Piferrer, 2001; Lin et al., 2012; Thuong et al., 2017; Alcántar-Vázquez, 2018; Vidal-López et al., 2019), in this study, we did not observe mortality and differences in food intake or growth among treatments and controls. According to Vidal-López et al. (2019), estrogens show no anabolic effect in most teleost. This agrees with that we observed in our study, since exposure to estrogen did not cause a significant reduction or increase in growth rate. Thus, 100 mg kg⁻¹ of E_2 may be used for this purpose in *L. macrocephalus*, since it did not cause productive losses.

It has been previously shown that the sexual differentiation in fish is regulated by sex steroid hormones (Singh, 2013; Pradhan and Olsson, 2015; Hoga et al., 2018) and indirectly by steroidogenic enzymes (Baroiller and D'Cotta, 2001; Tokarz et al., 2015; Di Rosa et al., 2016; Koyama et al., 2019). However, environmental factors (Hunter and Donaldson, 1983; Devlin and Nagahama, 2002; Piferrer et al., 2012; Díaz and Piferrer, 2015) and hormonal manipulations may intensify this process (Piferrer, 2001; Lin et al., 2012; Alcántar-Vázquez et al., 2015; Marín-Ramírez et al., 2016; Juárez-Juárez et al., 2017). In this concern, the most conventional route of ovarian development involves the transcription of the Cyp 19a1a gene and the production of the enzyme aromatase complex (P450arom). This enzyme is the main key in estrogen synthesis (Vernetti et al., 2013; Göppert et al., 2016), which is responsible for inducing and maintaining ovary development (Devlin and Nagahama, 2002; Nishimura and Tanaka, 2014; Lau et al., 2016). Thus, it is possible that diets supplemented with E₂ increase the circulating levels of this hormone in the treated fish, promoting the gonadal differentiation during the initial development of this species. In fact, this has been seen in several studies (Marín-Ramírez et al., 2016; Juárez-Juárez et al., 2017; Alcántar-Vázquez, 2018; Vidal-López et al., 2019) in which diets supplemented with E₂ promote fish gonadal differentiation.

The action of E_2 on gonadal differentiation in fish is well documented in the literature (Nagahama and Yamashita, 2008; Thuong et al., 2017; Alcántar-Vázquez, 2018; Vidal-López et al., 2019), and this steroid binds to specific nuclear receptors, which are three main subtypes in fish (α , β 1 and β 2) (Thomas et al., 2006). Such receptors are expressed at different sites and they also regulate ovarian development and growth (Pankhurst, 2016). In addition, during the early stages of growth, E_2 stimulates the first mitotic divisions of oogonia (Miura et al., 2002; Lubzens et al., 2017) and the synthesis of cortical alveoli in oocytes (Kwok et al., 2005; Luckenbach et al., 2013; Lubzens et al., 2017).

CONCLUSION

In this study, the use of diets supplemented with 100 mg kg⁻¹ of E_2 demonstrated to be an alternative to intensify the process of feminization in *L. macrocephalus*. Monosex fish production is a breakthrough for aquaculture, and several studies showed the efficiency of this technique to increase productivity. In this concern, here we have brought promising results for application of this methodology to *L. macrocephalus*, in which females present higher growth rates than males. However, future studies are needed to investigate the mechanism of action of E_2 in feminization in this species and to clearly determine the best dose and exact time for administration of diets enriched with E_2 to increase the rate of sexual inversion.

ACKNOWLEDGMENTS

Fundação de Amparo à Pesquisa do Estado de São Paulo (FAPESP) (grant #2010/08334-7 - doctoral scholarship).

REFERENCES

- Alcántar-Vázquez, J.P. 2018. Sex proportion in Nile tilapia Oreochromis niloticus fed estrogen mixtures: a case of paradoxical masculinization. Latin American Journal of Aquatic Research, 46(2): 337-345. http:// dx.doi.org/10.3856/vol46-issue2-fulltext-9.
- Alcántar-Vázquez, J.P.; Rueda-Curiel, P.; Calzada-Ruíz, D.; Antonio-Estrada, C.; Moreno-de la Torre, R. 2015. Feminization of Nile tilapia *Oreochromis niloticus* by estradiol-17β effects on growth, gonadal development, and body composition. Hidrobiológica, 25(2): 275-283.
- Baroiller, J.F.; D'Cotta, H. 2001. Environment and sex determination in farmed fish. Comparative Biochemistry and Physiology Part C: Toxicology & Pharmacology, 130(4): 399-409. http://dx.doi.org/10.1016/S1532-0456(01)00267-8.
- Budd, A.M.; Banh, Q.Q.; Domingos, J.A.; Jerry, D.R. 2015. Sex control in fish: approaches, challenges and opportunities for Aquaculture. Journal of Marine Science and Engineering, 3(2): 329-355. http://dx.doi. org/10.3390/jmse3020329.
- Capodifoglio, K.R.H.; Adriano, E.A.; Silva, M.R.M.; Maia, A.A.M. 2015. Supplementary data of *Henneguya leporinicola* (Myxozoa, Myxosporea) a parasite of *Leporinus macrocephalus* from fish farms in the state of São Paulo, Brazil. Acta Parasitologica, 60(3). http://dx.doi.org/10.1515/ ap-2015-0062.
- Cnaani, A.; Levavi-Sivan, B. 2009. Sexual development in fish, practical applications for aquaculture. Sexual Development, 3(2-3): 164-175. http://dx.doi.org/10.1159/000223080.
- Devlin, R.H.; Nagahama, Y. 2002. Sex determination and sex differentiation in fish: an overview of genetic, physiological, and environmental influences. Aquaculture, 208(3-4): 191-364. http://dx.doi.org/10.1016/ S0044-8486(02)00057-1.

- Di Rosa, V.; López-Olmeda, J.F.; Burguillo, A.; Frigato, E.; Bertolucci, C.; Piferrer, F.; Sánchez-Vázquez, F.J. 2016. Daily rhythms of the expression of key genes involved in steroidogenesis and gonadal function in zebrafish. PLoS One, 11(6): e0157716. http://dx.doi.org/10.1371/ journal.pone.0157716.
- Díaz, N.; Piferrer, F. 2015. Lasting effects of early exposure to temperature on the gonadal transcriptome at the time of sex differentiation in the European sea bass, a fish with mixed genetic and environmental sex determination. BMC Genomics, 16(1): 679. http://dx.doi.org/10.1186/ s12864-015-1862-0.
- Duke Energy International-Geração Paranapanema S/A. 2003. Peixes do rio Paranapanema. São Paulo: Horizonte Geográfico, 112p.
- Fernandino, J.I.; Hattori, R.S. 2019. Sex determination in Neotropical fish: Implications ranging from aquaculture technology to ecological assessment. General and Comparative Endocrinology, 273(1): 172-183. http://dx.doi.org/10.1016/j.ygcen.2018.07.002.
- Giamas, M.T.D.; Vermulm Junior, H. 2004. Levantamento da pesca profissional continental no Estado de São Paulo em 2001. In: São Paulo, Instituto de Pesca. Dados preliminares: bacia do Rio Paranapanema, Paraná e Grande. São Paulo: Instituto de Pesca. p. 1-10. (Série Relatórios Técnicos, 17). Available from: https://www.pesca.sp.gov.br/17_serreltec.pdf Access on: 7 July, 2020.
- Göppert, C.; Harris, R.M.; Theis, A.; Boila, A.; Hohl, S.; Rüegg, A.; Hofmann, H.A.; Salzburger, W.; Böhne, A. 2016. Inhibition of aromatase induces partial sex change in a cichlid fish: distinct functions for sex steroids in brains and gonads. Sexual Development, 10(2): 97-110. http://dx.doi. org/10.1159/000445463.
- Hashimoto, D.T.; Mendonça, F.F.; Senhorini, J.A.; Bortolozzi, J.; Oliveira, C.; Foresti, F.; Porto-Foresti, F. 2010. Identification of hybrids between Neotropical fish *Leporinus macrocephalus* and *Leporinus elongatus* by PCR-RFLP and multiplex-PCR: Tools for genetic monitoring in aquaculture. Aquaculture, 298(3-4): 346-349. http://dx.doi.org/10.1016/j. aquaculture.2009.11.015.
- Hoga, C.A.; Almeida, F.L.; Reyes, F.G.R. 2018. A review on the use of hormones in fish farming: analytical methods to determine their residues. CYTA: Journal of Food, 16(1): 679-691. http://dx.doi.org/ 10.1080/19476337.2018.1475423.
- Hunter, G.A.; Donaldson, E.M. 1983. Hormonal sex control and its application to fish culture. In: Hunter, G.A.; Donaldson, E.M. Fish physiology. New York: Academic Press. cap. 5. p. 223-303. http://dx.doi.org/10.1016/ \$1546-5098(08)60305-2.
- IBGE Instituto Brasileiro de Geografia e Estatística. 2018. Pesquisa da pecuária municipal 2018. Available from: https://sidra.ibge.gov.br/ pesquisa/ppm/quadros/brasil/2018> Access on: 6 May, 2020.
- Juárez-Juárez, V.; Alcántar-Vázquez, J.P.; Antonio-Estrada, C.; Marín-Ramírez, J.A.; Moreno-de la Torre, R. 2017. Feminization by 17*a*-ethinylestradiol of the progeny of XY-female Nile tilapia (*Oreochromis niloticus*). Effects on growth, condition factor and gonadosomatic index. Turkish Journal of Fisheries and Aquatic Sciences, 17(3): 599-607. http:// dx.doi.org/10.4194/1303-2712-v17_3_16.

- Koyama, T.; Nakamoto, M.; Morishima, K.; Yamashita, R.; Yamashita, T.; Sasaki, K.; Kuruma, Y.; Mizuno, N.; Suzuki, M.; Okada, Y.; Ieda, R.; Uchino, T.; Tasumi, S.; Hosoya, S.; Uno, S.; Koyama, J.; Toyoda, A.; Kikuchi, K.; Sakamoto, T. 2019. A SNP in a steroidogenic enzyme is associated with phenotypic sex in seriola fishes. Current Biology, 29(11): 1901-1909. http://dx.doi.org/10.1016/j.cub.2019.04.069.
- Kwok, H.; So, W.K.; Wang, Y.; Ge, W. 2005. Zebrafish gonadotropins and their receptors: I. cloning and characterization of zebrafish folliclestimulating hormone and luteinizing hormone receptors – evidence for their distinct functions in follicle development. Biology of Reproduction, 72(6): 1370-1381. http://dx.doi.org/10.1095/biolreprod.104.038190.
- Lau, E.S.; Zhang, Z.W.; Qin, M.; Ge, W. 2016. Knockout of zebrafish ovarian aromatase gene (cyp19a1a) by TALEN and CRISPR/Cas9 leads to allmale off spring due to failed ovarian differentiation. Scientific Reports, 6(1): 37357. http://dx.doi.org/10.1038/srep37357.
- Lin, S.; Benfey, T.J.; Martin-Robichaud, M.D. 2012. Hormonal sex reversal in Atlantic cod, *Gadus morhua*. Aquaculture, 364–365: 192-197. http:// dx.doi.org/10.1016/j.aquaculture.2012.08.023.
- Lubzens, E.; Bobe, J.; Young, G.; Sullivan, C.V. 2017. Maternal investment in fish oocytes and eggs: The molecular cargo and its contributions to fertility and early development. Aquaculture, 472: 107-143. http:// dx.doi.org/10.1016/j.aquaculture.2016.10.029.
- Luckenbach, J.A.; Yamamoto, Y.; Guzmán, J.M.; Swanson, P. 2013. Identification of ovarian genes regulated by follicle-stimulating hormone (Fsh) in vitro during early secondary oocyte growth in coho salmon. Molecular and Cellular Endocrinology, 366(1): 38-52. http://dx.doi.org/10.1016/j. mce.2012.11.015.
- Marín-Ramírez, J.A.; Alcántar-Vázquez, J.P.; Antonio-Estrada, C.; Moreno-de la Torre, R.; Calzada-Ruiz, D. 2016. Feminization of Nile tilapia *Oreochromis niloticus* (L.) by diethylstilbestrol: growth and gonadosomatic index. Ecosistemas y Recursos Agropecuarios, 3(7): 51-61.
- Miura, T.; Miura, C.; Konda, Y.; Yamauchi, K. 2002. Spermatogenesispreventing substance in Japanese eel. Development, 129(11): 2689-2697.
- Morelli, K.A.; Revaldaves, E.; Oliveira, C.; Foresti, F. 2007. Isolation and characterization of eight microsatellite loci in *Leporinus macrocephalus* (Characiformes: Anostomidae) and cross-species amplification. Molecular Ecology Notes, 7(1): 32-34. http://dx.doi. org/10.1111/j.1471-8286.2006.01484.x.
- Muñoz, M.E.; Batlouni, S.R.; Vicentini, I.B.F.; Vicentini, C.A. 2011. Testicular structure and description of the seminal pathway in *Leporinus macrocephalus* (Anostomidae, Teleostei). Micron, 42(8): 892-897. http://dx.doi.org/10.1016/j.micron.2011.06.008.
- Nagahama, Y.; Yamashita, M. 2008. Regulation of oocyte maturation in fish. Development, Growth & Differentiation, 50(1): 195-219. http://dx.doi. org/10.1111/j.1440-169X.2008.01019.x.
- Nishimura, T.; Tanaka, M. 2014. Gonadal Development in Fish. Sexual Development, 8(5): 252-261. http://dx.doi.org/10.1159/000364924.
- Örn, S.; Holbech, H.; Norrgren, L. 2016. Sexual disruption in zebrafish (*Danio rerio*) exposed to mixtures of 17α-ethinylestradiol and 17β-trenbolone. Environmental Toxicology and Pharmacology, 41: 225-231. http:// dx.doi.org/10.1016/j.etap.2015.12.010.
- Pankhurst, N.W. 2016. Reproduction and development. biology of stress in fish. Fish Physiology, 35: 295-331. http://dx.doi.org/10.1016/ B978-0-12-802728-8.00008-4.

- Peixe BR. 2019. Anuário Peixe BR da piscicultura 2019. São Paulo: Associação Brasileira de Piscicultura, 2019. 148p.
- Pereira, T.S.B.; Boscolo, C.N.P.; Moreira, R.G.; Batlouni, S.R. 2017. The use of mGnRHa provokes ovulation but not viable embryos in *Leporinus macrocephalus*. Aquaculture International, 25(2): 515-529. http:// dx.doi.org/10.1007/s10499-016-0049-2.
- Pereira, T.S.B.; Boscolo, C.N.P.; Silva, D.G.H.; Batlouni, S.R.; Schlenk, D.; Almeida, E.D. 2015. Anti-androgenic activities of diuron and its metabolites in male Nile tilapia (*Oreochromis niloticus*). Aquatic Toxicology, 164: 10-15. http://dx.doi.org/10.1016/j.aquatox.2015.04.013.
- Petrere Junior, M.; Agostinho, A.A.; Okada, E.K.; Julio Junior, H.F. 2002. Review of the fisheries in the Brazilian portion of the Paraná/Pantanal basin. In: Cowx, I.G. (Ed.). Management and ecology of lake and reservoir fisheries. Oxford: Fishing News Books, p. 123-143.
- Piferrer, F. 2001. Endocrine sex control strategies for the feminization of teleost fish. Aquaculture, 197(1-4): 229-281. http://dx.doi.org/10.1016/ S0044-8486(01)00589-0.
- Piferrer, F.; Ribas, L.; Díaz, N. 2012. Genomic approaches to study genetic and environmental influences on fish sex determination and differentiation. Marine Biotechnology, 14(5): 591-604. http://dx.doi.org/10.1007/ s10126-012-9445-4.
- Pradhan, A.; Olsson, P.E. 2015. Zebrafish sexual behavior: role of sex steroid hormones and prostaglandins. Behavioral and Brain Functions, 11(1): 23. http://dx.doi.org/10.1186/s12993-015-0068-6.
- Reidel, A.; Oliveira, L.G.; Piana, P.A.; Lemainski, D.; Bombardelli, R.A.; Boscolo, W.R. 2004. Avaliação de rendimento e características morfometricas do curimbatá *Prochilodus lineatus* (VALENCIENNES, 1836), e do piavuçu *Leporinus macrochephalus* (GARAVELLO & BRITSKI, 1988) machos e fêmeas. Revista Varia Scientia, 4(8): 71-78.
- Riffel, A.P.K.; Garcia, L.O.; Finamor, I.A.; Saccol, E.M.H.; Meira, M.; Kolberg, C.; Horst, A.; Partata, W.; Llesuy, S.; Baldisserotto, B.; Pavanato, M.A. 2012. Redox profile in liver of *Leporinus macrocephalus* exposed to different dissolved oxygen levels. Fish Physiology and Biochemistry, 38(3): 797-805. http://dx.doi.org/10.1007/s10695-011-9563-3.
- Singh, A.K. 2013. Introduction of modern endocrine techniques for the production of monosex population of fishes. General and Comparative Endocrinology, 181: 146-155. http://dx.doi.org/10.1016/j.ygcen.2012.08.027.
- Soares, C.M.; Hayashi, C.; Furuya, V.R.B.; Furuya, W.M.; Galdioli, E.M. 2000. Substituição parcial e total da proteína do farelo de soja pela proteína do farelo de canola na alimentação de alevinos de piavuçu (*Leporinus macrocephalus*, L.). Revista Brasileira de Zootecnia, 29(1): 15-22. http://dx.doi.org/10.1590/S1516-35982000000100003.
- Thomas, P.; Dressing, G.; Pang, Y.; Berg, H.; Tubbs, C.; Benninghoff, A.; Doughty, K. 2006. Progestin, estrogen and androgen G-protein coupled receptors in fish gonads. Steroids, 71(4): 310-316. http://dx.doi. org/10.1016/j.steroids.2005.09.015.
- Thuong, N.P.; Sung, Y.Y.; Ambak, M.A.; Abol-Munafi, A.B. 2017. The hormone 17 β-estradiol promotes feminization of juveniles protandrous hermaphrodite false clownfish (*Amphiprion ocellaris*). Marine and Freshwater Behaviour and Physiology, 50(3): 195-204. http://dx.doi. org/10.1080/10236244.2017.1361788.
- Tokarz, J.; Möller, G.; Hrabě de Angelis, M.; Adamski, J. 2015. Steroids in teleost fishes: a functional point of view. Steroids, 103: 123-144. http:// dx.doi.org/10.1016/j.steroids.2015.06.011.

- Vernetti, C.H.M.M.; Rodrigues, M.D.N.; Gutierrez, H.J.P.; Calabuig, C.P.; Moreira, C.G.A.; Nlewadim, A.A.; Moreira, H.L.M. 2013. Genes involved in sex determination and the influence of temperature during the sexual differentiation process in fish: a review. African Journal of Biotechnology, 12(17): 2129-2146. http://dx.doi.org/10.5897/AJB12.1155.
- Vidal-López, J.M.; Contreras-Sánchez, W.M.; Hernández-Franyutti, A.; Contreras-García, M.J.; Uribe-Aranzábal, M.C. 2019. Functional feminization of the Mexican snook (*Centropomus poeyi*) using 17β-estradiol in the diet. Latin American Journal of Aquatic Research, 47(2): 240-250. http://dx.doi.org/10.3856/vol47-issue2-fulltext-4.